Oncostatin M suppresses browning of white adipocytes via gp130-STAT3 signaling

van Krieken, Pim P ; Odermatt, Timothy S ; Borsigova, Marcela ; Blüher, Matthias ; Wueest, Stephan ; Konrad, Daniel

Abstract: Objective Obesity is associated with low-grade adipose tissue inflammation and locally elevated levels of several glycoprotein 130 (gp130) cytokines. The conversion of white into brown-like adipocytes (browning) may increase energy expenditure and revert the positive energy balance that underlies obesity. Although different gp130 cytokines and their downstream targets were shown to regulate expression of the key browning marker uncoupling protein 1 (Ucp1), it remains largely unknown how this contributes to the development and maintenance of obesity. Herein, we aim to study the role of gp130 cytokine signaling in white adipose tissue (WAT) browning in the obese state. Methods Protein and gene expression levels of UCP1 and other thermogenic markers were assessed in a subcutaneous adipocyte cell line, adipose tissue depots from control or adipocyte-specific gp130 knockout (gp130Δadipo) mice fed either chow or a high-fat diet (HFD), or subcutaneous WAT biopsies from a human cohort of lean and obese subjects. WAT browning was modelled in vitro by exposing mature adipocytes to isoproterenol subsequent to stimulation with gp130 cytokines. ERK and JAK-STAT signaling were blocked using the inhibitors U0126 and Tofacitinib, respectively. Results Inguinal WAT of HFD-fed gp130Δadipo mice exhibited significantly elevated levels of UCP1 and other browning markers such as Cidea and Pgc-1 . In vitro, treatment with the gp130 cytokine oncostatin M (OSM) lowered isoproterenol-induced UCP1 protein and gene expression levels in a dose-dependent manner. Mechanistically, OSM mediated the inhibition of Ucp1 via the JAK-STAT but not the ERK pathway. In line with mouse data, OSM gene expression in human WAT positively correlated with BMI (r=0.284, p=0.021, n=66) and negatively with UCP1 expression (r=-0.413, p<0.001, n=66). Conclusions Our data support the notion that OSM negatively regulates thermogenesis in WAT and, thus, may be an attractive target to treat obesity.

DOI: https://doi.org/10.1016/j.molmet.2021.101341

van Krieken, Pim P; Odermatt, Timothy S; Borsigova, Marcela; Blüher, Matthias; Wueest, Stephan; Konrad, Daniel (2021). Oncostatin M suppresses browning of white adipocytes via gp130-STAT3 signaling. Molecular Metabolism, 54:101341.
DOI: https://doi.org/10.1016/j.molmet.2021.101341

Oncostatin M suppresses browning of white adipocytes via gp130-STAT3 signaling

Pim P. van Krieken, Timothy S. Odermatt, Marcela Borsigova, Matthias Blüher, Stephan Wueest, Daniel Konrad

PII: S2212-8778(21)00188-5

DOI: https://doi.org/10.1016/j.molmet.2021.101341

Reference: MOLMET 101341

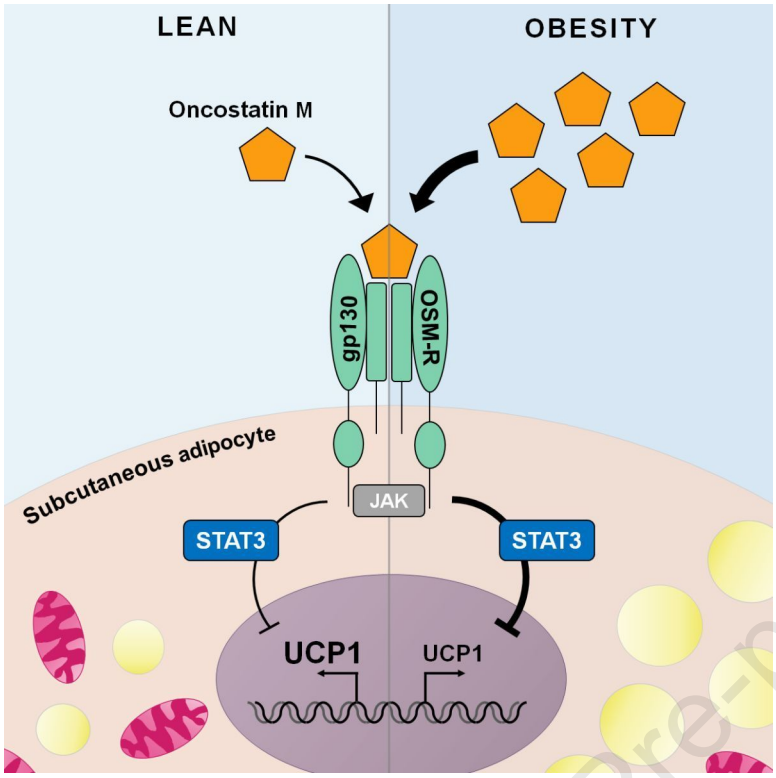
To appear in: *Molecular Metabolism*

Received Date: 2 June 2021

Revised Date: 13 September 2021

Accepted Date: 13 September 2021

Please cite this article as: van Krieken PP, Odermatt TS, Borsigova M, Blüher M, Wueest S, Konrad D, Oncostatin M suppresses browning of white adipocytes via gp130-STAT3 signaling, *Molecular Metabolism*, https://doi.org/10.1016/j.molmet.2021.101341.



Oncostatin M suppresses browning of white adipocytes via gp130-STAT3 signaling

Pim P. van Krieken[1,2], Timothy S. Odermatt[1,2,3], Marcela Borsigova[1,2], Matthias Blüher[4, 5], Stephan Wueest[1,2,*], Daniel Konrad[1,2,3,*]

[1]Division of Pediatric Endocrinology and Diabetology and [2]Children's Research Center, University Children's Hospital, University of Zurich, CH-8032 Zurich, Switzerland

[3]Zurich Center for Integrative Human Physiology, University of Zurich, CH-8057 Zurich, Switzerland

[4]Department of Medicine, Endocrinology and Diabetes, University of Leipzig, D-04103 Germany

[5]Helmholtz Institute for Metabolic, Obesity and Vascular Research (HI-MAG) of the Helmholtz Zentrum München at the University of Leipzig and University Hospital Leipzig, Germany

*These authors contributed equally to this work

Correspondence to:

Daniel Konrad, MD PhD

University Children's Hospital

Department of Endocrinology and Diabetology

Steinwiesstrasse 75

CH-8032 Zurich

Tel: ++41-44-266 7966; Fax: ++41-44-266 7983

Email: daniel.konrad@kispi.uzh.ch

Abstract

Objective Obesity is associated with low-grade adipose tissue inflammation and locally elevated levels of several glycoprotein 130 (gp130) cytokines. The conversion of white into brown-like adipocytes (browning) may increase energy expenditure and revert the positive energy balance that underlies obesity. Although different gp130 cytokines and their downstream targets were shown to regulate expression of the key browning marker *uncoupling protein 1* (*Ucp1*), it remains largely unknown how this contributes to the development and maintenance of obesity. Herein, we aim to study the role of gp130 cytokine signaling in white adipose tissue (WAT) browning in the obese state.

Methods Protein and gene expression levels of UCP1 and other thermogenic markers were assessed in a subcutaneous adipocyte cell line, adipose tissue depots from control or adipocyte-specific gp130 knockout ($gp130^{\Delta adipo}$) mice fed either chow or a high-fat diet (HFD), or subcutaneous WAT biopsies from a human cohort of lean and obese subjects. WAT browning was modelled *in vitro* by exposing mature adipocytes to isoproterenol subsequent to stimulation with gp130 cytokines. ERK and JAK-STAT signaling were blocked using the inhibitors U0126 and Tofacitinib, respectively.

Results Inguinal WAT of HFD-fed $gp130^{\Delta adipo}$ mice exhibited significantly elevated levels of UCP1 and other browning markers such as Cidea and Pgc-1α. *In vitro*, treatment with the gp130 cytokine oncostatin M (OSM) lowered isoproterenol-induced UCP1 protein and gene expression levels in a dose-dependent manner. Mechanistically, OSM mediated the inhibition of *Ucp1* via the JAK-STAT but not the ERK pathway. In line with mouse data, *OSM* gene expression in human WAT positively correlated with BMI ($r=0.284$, $p=0.021$, $n=66$) and negatively with *UCP1* expression ($r=-0.413$, $p<0.001$, $n=66$).

Conclusions Our data support the notion that OSM negatively regulates thermogenesis in WAT and, thus, may be an attractive target to treat obesity.

Keywords:

Browning, high fat diet, obesity, oncostatin M, UCP1, white adipose tissue

Abbreviations:

BAT, brown adipose tissue; CM, complete medium; epi, epididymal; ERK, extracellular signal-regulated kinase; gp130, glycoprotein 130; $gp130^{\Delta adipo}$, adipocyte-specific glycoprotein 130 knockout; HFD, high-fat diet; ing, inguinal; JAK, janus kinase; OSM, oncostatin M; $OSMR^{\Delta adipo}$, adipocyte-specific oncostatin M receptor knockout; SOCS3, suppressor of cytokine signaling 3; STAT, signal transducer and activator of transcription; SVF, stromal vascular fraction; TBS-T, tris-buffered saline with 0.1% Tween; UCP1, uncoupling protein 1; WAT, white adipose tissue.

1. Introduction

Obesity manifests upon a sustained positive energy balance throughout which excess energy is stored in white adipose tissue (WAT) depots. The key to reversing obesity therefore lies in attaining a negative energy balance. Brown adipocytes can dissipate energy through non-shivering thermogenesis, an essential process to maintain core temperature requiring uncoupling protein 1 (UCP1). Besides cold exposure-mediated stimulation of β-adrenergic receptors, which is the main physiological stimulus to activate brown adipose tissue (BAT), a large number of alternative stimuli including exercise and specific diets have been reported (1; 2). However, since obese individuals seem to have rather small amounts of functional BAT (3-5), it may be more promising to target their WAT depots instead. Namely, WAT can be harnessed to burn rather than store energy by conversion of white into brown-like adipocytes (browning). Consequently, the focus of obesity research on WAT browning has gained substantial momentum over the past decade (6). Despite huge efforts, only a small number of drugs with proven or presumed effects on WAT browning are presently available in clinic (7), making the search for better alternatives imperative.

The expansion of adipose tissue depots is paralleled by an invasion of immune cells that concomitantly leads to a state of low-grade inflammation (8). Immune cells communicate with adipocytes and modulate their metabolic functions via the secretion of pro-inflammatory cytokines. The glycoprotein 130 (gp130) cytokine family comprises a main subgroup of such cytokines that consists of interleukin (IL)-6, IL-11, IL-27, IL-35, cardiotrophin-1, cardiotrophin-like cytokine, ciliary neurotrophic factor, leukemia inhibitory factor and oncostatin M (OSM). Although each of these cytokines has a unique receptor on the surface of the adipocyte to mediate intracellular signaling, each receptor consists

of at least one gp130 subunit making gp130 the common signal transducer for all family members (9; 10). The two key intracellular signaling cascades activated by all gp130 cytokines are the extracellular signal-regulated kinase (ERK) and Janus kinase (JAK)-signal transducer and activator of transcription (STAT) pathways (11). As such, gp130 cytokines can influence a plethora of metabolic processes. For instance, making use of mice that lacked the gp130 protein specifically in adipocytes ($gp130^{\Delta adipo}$ mice), we previously unveiled that gp130 cytokine signaling regulates free-fatty acid and leptin release in WAT, thereby indirectly affecting insulin release from pancreatic islets (12; 13). Interestingly, several signaling molecules upstream as well as downstream of the gp130 signaling complex have been related to WAT browning. For example, activation of both the ERK and the JAK-STAT pathway in adipocytes was associated with either positive or negative changes in WAT browning (14-17). Furthermore, the gp130 cytokines IL-6 and OSM were linked to more and less UCP1 expression in WAT, respectively (18-20). Although they are both elevated in WAT under obesity (20; 21), their role in WAT browning in the latter condition remains largely unknown. Herein, we explored the role of gp130 signaling in WAT browning in the obese state using previously characterized $gp130^{\Delta adipo}$ mice. Moreover, by performing experiments in cultured white adipocytes and in human WAT, we aimed to shed light on the opposing claims of up- and downstream players of gp130 in this process.

2. Material and Methods

2.1 Human samples

We included 159 individuals with a wide range of BMI (19-73 kg/m^2), type 2 diabetes (n=84) and normal glucose metabolism (n=75). Paired abdominal subcutaneous adipose tissue biopsies were taken during elective sleeve gastrectomy, Roux-en-Y gastric bypass, hernia or cholecystectomy surgeries and processed as previously described (22). The study was approved by the Ethics Committee of the University of Leipzig (approval no: 159-12-21052012), and performed in accordance to the declaration of Helsinki. All subjects gave written informed consent before taking part in this study. RNA from adipose tissue was extracted by using RNeasy Lipid tissue Mini Kit (Qiagen, Hilden, Germany). Quantity and integrity of RNA was monitored with NanoVue plus Spectrophotometer (GE Healthcare, Freiburg, Germany). 1 µg total RNA from subcutaneous adipose tissue was reverse-transcribed with standard reagents (Life technologies, Darmstadt, Germany). cDNA was then processed for TaqMan probe-based quantitative real-time polymerase chain reaction (qPCR) using the QuantStudio 6 Flex Real-Time PCR System (Life technologies, Darmstadt, Germany). Expression of *OSM*, *UCP1*, and *IL-6* were calculated by standard curve method and normalized to the expression of *hypoxanthine guanine phosphoribosyltransferase 1* (*HPRT1*) as a housekeeping gene. The probes (Life technologies, Darmstadt, Germany) for *OSM* (Hs00171165_m1), *UCP1* (Hs00222453_m1), *IL-6* (Hs00985639_m1) and *HPRT1* (Hs01003267_m1) span exon-exon boundaries to improve the specificity of the qPCR. For analysis, only subjects in whom all three genes were detected (n=66) were included.

2.2 Mouse samples

To obtain gp130$^{\Delta adipo}$ mice, gp130$^{F/F}$ mice (23) were crossed with Adipoq-Cre mice (The Jackson Laboratory, Bar Harbor, ME, USA) as described (12). Littermate mice with floxed gp130 but absent Cre-recombinase (Cre) expression were used as controls (gp130$^{F/F}$). Every animal was considered an experimental unit. Animals were housed in groups of 2-5 in a specific pathogen-free, temperature-controlled environment (±22°C) with a 12 h light/12 h dark cycle and crinkles as bedding material. Mice received standard chow diet (ProvimiKliba, Kaiseraugst, Switzerland) or were switched to HFD (D12331; Research Diets, New Brunswick, NJ, USA) at the age of 6 weeks. Animals were allocated to diets based on body weight (similar mean +/- SD in starting body weight between the groups). Water was provided ad libitum. Experimenters were not blinded to the identity of a specific mouse. Adipose tissue samples were harvested from 18-week-old male mice after a 5-hour fasting period. Samples were snap-frozen and stored at -80°C. Experiments were performed according to Swiss animal protection laws and approved by the cantonal veterinary office in Zurich, Switzerland.

2.3 Cell culture and treatments

Previously characterized murine subcutaneous white pre-adipocytes (24) or 3T3-L1 cells were plated in 12- or 24-well plates coated with 0.1% wt/vol. gelatin (Bio-Rad 170-6537). Cells were maintained in complete medium (CM) consisting of DMEM (25 mmol/l glucose) supplemented with 10% vol./vol. fetal bovine serum and 1% vol./vol. penicillin/streptomycin (all from Invitrogen, Basel, Switzerland) that was changed every 2-3 days. Differentiation towards mature adipocytes started two days after reaching confluency. The first 72 hours, CM was supplemented with 3-isobutyl-1-methylxanthine

(500 µmol/l), dexamethasone (1 µmol/l), insulin (1.7 µmol/l) and rosiglitazone (1 µmol/l). The next 2 days cells were cultured in CM with insulin (0.5 µmol/l). Experiments with subcutaneous adipocytes were commenced after another 2 days in CM without further supplements. Differentiation of 3T3-L1 cells, cultivation of RAW 264.7 cells, and isolation of adipocytes and stromal vascular fraction (SVF) was performed as previously described (25; 26).

Fully differentiated adipocytes were treated with vehicle or various concentrations of OSM (495-MO, R&D systems, Zug, Switzerland), IL-6 (AG-40B-0108, AdipoGen, Liestal, Switzerland) or leptin (498-OB, R&D systems, Zug, Switzerland) for up to 24 hours as indicated in the figure legends. Browning was induced by 3 or 6 hours treatment with 1 or 10 µmol/l isoproterenol as indicated. For inhibitor experiments, cells were pre-treated for 1 hour with 50 µmol/l U0126 (ab120241, Abcam, Cambridge, UK), 2 µmol/l Tofacitinib (CP-690550, LuBioScience GmbH, Zurich, Switzerland), or a vehicle. Isolated SVF cells or cultured RAW cells were treated with 10 ng/ml lipopolysaccharides (LPS; Sigma-Aldrich, Buchs, Switzerland) for 6 hours. 3T3-L1 cells were treated with 100 nM human insulin (Actrapid, Novo Nordisk Pharma AG, Zurich, Switzerland) for 10 minutes. Cells were harvested after a brief wash in ice-cold PBS. Cell culture data represent at least two biological replicates and each condition was generally performed in duplicates. Cells were not tested for mycoplasma contamination. Experimenters were not blinded to the identity of a specific sample.

2.4 Protein lysis and western blot analysis

Tissues or cells were lysed in a buffer containing 150 mmol/l NaCl, 50 mmol/l Tris-HCl (pH 7.5), 1 mmol/l EGTA, 1% vol./vol. NP-40, 0.25% wt/vol. sodium deoxycholate, 1

mmol/l sodium pyrophosphate, 1 mmol/l sodium vanadate, 1 mmol/l NaF, 10 mmol/l sodium β-glycerolphosphate, 0.2 mmol/l PMSF and 0.1% vol./vol. protease inhibitor cocktail (Sigma-Aldrich, Buchs, Switzerland). Protein concentrations were determined with a BCA assay (Pierce, Rockford, IL, USA). Equal amounts of protein were resolved using SDS-PAGE and electro-transferred onto 0.2 µm nitrocellulose membranes (BioRad, Reinach, Switzerland). Equal loading of proteins was verified by Ponceau S staining. Blots were blocked with 5% wt/vol. dry fat milk dissolved in tris-buffered saline (150 mmol/l NaCl, 50 mmol/l Tris-HCl) containing 0.1% vol./vol. Tween (TBS-T). Membranes were incubated overnight at 4°C with primary antibody diluted in TBS-T supplemented with 5% wt/vol. bovine serum albumin and 0.02% wt/vol. sodium azide followed by 1 hour incubation at room temperature with a secondary antibody diluted in TBS-T with 5% wt/vol. dry fat milk. Membranes were developed using the ECL system and antibody-antigen complexes were detected with a ChemiDoc™ MP Imaging System (Biorad, Reinach, Switzerland). Primary antibodies were used at a dilution of 1:1000 unless specified and include: UCP1, PA1-24894 (Thermofisher Scientific, Waltham, MA, USA); pAkt (Thr308), #13038; Akt, #9272; pERK1/2, #9101; ERK1/2, #9211; pSTAT3, #9145; STAT3, #9132 (Cell Signaling, Danvers, MA, USA); PPARγ, sc-7196 (1:200, Santa Cruz Biotechnology, Santa Cruz, CA, USA); and Actin, MAB1501 (Millipore, Darmstadt, Germany). Molecular weight/pattern of bands was compared to existing literature when possible. Details of antibodies used such as data sheets, citations and reviews can be found via antibody above listed catalogue numbers. Secondary antibodies, diluted 1:5000, are goat-anti-rabbit, ab6721 and mouse-anti-rabbit, ab6789 (Abcam, Cambridge, UK).

2.5 RNA extraction and RT-qPCR analysis

Total RNA was extracted from homogenized tissues with the NucleoSpin® RNA Set for NucleoZOL or from cells with NucleoSpin® RNA isolation kit (Macherey-Nagel, Düren, Germany). RNA concentration was determined with NanoDrop® (ThermoFisher Scientific, Waltham, MA, USA). Equal amounts of RNA (50-500 ng) were reverse-transcribed with the GoScript™ Reverse Transcription System (Promega, Madison, WI, USA) and cDNA was amplified by real-time PCR using the following probes/primers: *Ucp1*, Mm0124486_m1; *Cidea*, Mm00432554_m1; *Ppargc1a*, Mm01208835_m1; *Prdm16*, Mm00712556_m1; *Dio2*, Mm00515664_m1; *Pparg*, Mm00440940_m1; *Ppara*, Mm00627559_m1; *Tbx1*, Mm00448949_m1; *Tmem26*, Mm01173641_m1; *Osm*, Mm01193966_m1; *gp130 (Il6st)*, Mm00439665_m1; 18s, 4352930 (Applied Biosystems, Rotkreuz, Switzerland). Autotaxin (*Enpp2*), Fw: GACCCTAAAGCCATTATTGCTAA, Rv: GGGAAGGTGCTGTTTCATGT (27); Probe: AAACCAGATCAGCACTTTAAGCC (Microsynth, Balgach, Switzerland). Relative gene expression values were obtained after normalization to 18s using the $2^{-\Delta\Delta Ct}$ method. Outliers defined as values exceeding 2 SD from the mean were excluded.

2.6 Hematoxylin-eosin and Oil-Red-O staining

Inguinal snap-frozen tissues were fixed overnight in 4% buffered formalin, dehydrated and embedded in paraffin blocks. Four micrometer thick sections were cut and stained for hematoxylin and eosin. For Oil-Red-O staining, cells were fixed in 4% wt/vol. paraformaldehyde solution (AppliChem, Darmstadt, Germany) for 30 minutes on ice, rinsed in dH_2O and dehydrated in 60% vol./vol. isoproterenol. Cells were stained with a solution of 0.3% wt/vol. Oil-Red-O (O0625; Sigma-Aldrich, Buchs, Switzerland)

dissolved in 100% isoproterenol that was freshly diluted to 60% vol./vol. in dH_2O. After 5-10 minutes, cells were rinsed twice in dH_2O. Tissue slices or cells were imaged using an Axio Observer microscope platform (Zeiss, Jena, Germany). Tile scans were analyzed with ImageJ software to calculate the Oil-Red-O positive signal per well.

2.7 Data analysis

Date are presented as means ± SE. Statistics were calculated using GraphPad Prism 8.00 (GraphPad Software, San Diego, CA, USA) by means of a paired or unpaired two-tailed Student's t-test (for normally distributed data), Mann-Whitney test (for not normally distributed data), one-way ANOVA with Tukey multiple comparisons correction, or linear regression analysis. A p-value < 0.05 was considered statistically significant. Power calculation analysis was not performed. Sample size was determined based on previous experiments performed in our laboratory.

2.8 Data and resource availability

The data supporting the findings of this study are available within the article, its Supplementary Information files or from the corresponding author upon reasonable request. No applicable resources were generated or analyzed during the current study.

3. Results

3.1 Increased browning in ingWAT of HFD-fed gp130$^{\Delta adipo}$ mice

To test the hypothesis that gp130 plays a role in WAT browning, we examined the levels of UCP1 in white fat depots obtained from our previously characterized chow and high-fat diet (HFD)-fed sedentary and exercised gp130$^{\Delta adipo}$ mice (12; 13; 28). The level of UCP1 was unchanged in inguinal (ing) WAT of chow fed mice and undetectable in the less browning-prone epididymal (epi) WAT irrespective of diet and genotype (Supplementary Fig. 1B-D). However, UCP1 protein levels were significantly elevated in ingWAT of HFD-fed gp130$^{\Delta adipo}$ mice compared to littermate controls (Fig. 1A-B). Similarly, UCP1 protein levels and number of multi-locular cells were increased in ingWAT of exercised HFD-fed gp130$^{\Delta adipo}$ mice, as illustrated by Western blot and histological analysis, respectively (Supplementary Fig. 1E-F). As expected, 12 weeks of HFD feeding markedly lowered *Ucp1* mRNA and UCP1 protein levels in ingWAT of control mice (Supplementary Fig. 1G-H). The above data infer that HFD-fed gp130$^{\Delta adipo}$ mice are partly protected from such decline in UCP1. To corroborate the latter observation we performed complementary gene expression analysis of ingWAT depots from HFD-fed mice and found increased *Ucp1* mRNA levels in knockout compared to control mice. Moreover, the transcripts of additional browning-related markers including *Cidea*, *Pgc-1α* (*Ppargc1a*) and *Tbx1* were significantly elevated (Fig. 1C). Of note, no significant difference in UCP1 protein level was detected when comparing BAT depots of gp130$^{\Delta adipo}$ and gp130$^{F/F}$ mice fed either regular chow (Supplementary Fig. 1A) or HFD (28). Together, these results indicate that while gp130 depletion may not affect thermogenic potential of classical BAT, it protects from the HFD-induced reduction in WAT browning.

Although we previously reported that neither body weight nor energy expenditure were significantly different between HFD-fed gp130$^{\Delta adipo}$ and gp130$^{F/F}$ mice (12; 28), we hitherto analyzed 17 HFD-fed litters of both genotypes (n=43 mice for gp130$^{F/F}$; n=35 mice for gp130$^{\Delta adipo}$) and observed that the average body weight per litter was significantly reduced in gp130$^{\Delta adipo}$ mice (-4.3±1.7% body weight; p=0.02). Importantly, body weight negatively correlated with *Ucp1* expression in ingWAT of HFD-fed mice (Fig. 1D), suggesting that the observed increase in browning contributed to reduced weight gain.

We next assessed levels of phosphorylated ERK1/2 (p-ERK1/2) and STAT3 (p-STAT3), both representing key proteins in the signaling cascades downstream of the gp130 receptor complex (11). Levels of p-ERK1/2 and p-STAT3 were distributed rather heterogeneously in ingWAT of HFD-fed gp130$^{\Delta adipo}$ and gp130$^{F/F}$ mice (Fig. 1A). Correlative analysis showed that p-STAT3 but not p-ERK1/2 negatively associated with UCP1 (Fig. 1E-F), suggesting a possible involvement of the STAT3 signaling pathway in the regulation of *Ucp1* transcription.

3.2 OSM dose-dependently lowers isoproterenol-induced UCP1 levels in cultured adipocytes

As outlined above, the gp130 cytokine family members IL-6 and OSM are elevated in obesity and may impact on UCP1 levels (18-20; 29). Moreover, the adipokine leptin, secretion and circulating levels of which were reduced in HFD-fed gp130$^{\Delta adipo}$ mice (12), may modulate WAT browning (30-32). Expression of the gp130 downstream mediator and potential suppressor of WAT browning autotaxin (gene name *Enpp2*) (33; 34) was not different between both genotypes in ingWAT (Fig. 1C) and, thus, seemed unlikely to contribute to elevated UCP1 in ingWAT of HFD-fed gp130$^{\Delta adipo}$ mice. The impact of the

three other candidates on browning was evaluated using a mouse-derived subcutaneous white adipocyte cell line. Basal *Ucp1* mRNA and UCP1 protein levels were low and remained unchanged upon stimulation with either OSM or IL-6 (Supplementary Fig. 2A, 2B). The subsequent induction of browning by the β_3-adrenoreceptor agonist isoproterenol strongly induced UCP1 expression, as expected. Intriguingly, whilst no change in isoproterenol-mediated induction of UCP1 level was noted when adipocytes were stimulated with IL-6 or leptin, a dose-dependent reduction was observed upon incubation with OSM (Fig. 2A-B; Supplementary Fig. 2C). OSM elicited comparable effects in a second adipocyte cell line, 3T3-L1 cells, by dose-dependently lowering *Ucp1* gene expression induced by isoproterenol treatment (Fig. 2C). These results indicate that OSM suppresses isoproterenol-induced UCP1 levels in cultured white adipocytes and, thus, suggest that elevated WAT browning in HFD-fed gp130$^{\Delta adipo}$ mice can at least in part be attributed to their lack in OSM signaling.

Within the adipose tissue depot, immune cells are the main source of OSM and may be responsible for its increased levels under obesity (20; 35). In line, we found higher *Osm* gene expression in the immune-cell containing SVF compared to adipocytes isolated from wild-type mice. Moreover, *Osm* expression tended to be elevated in samples from HFD-fed compared to chow-fed mice (Supplementary Fig. 3A). In contrast, no major differences were observed in *gp130* mRNA levels between the two cell fractions irrespective of diet (Supplementary Fig. 3B). We hypothesized that an inflammatory milieu as commonly present in fat depots under obesity may trigger OSM production in local immune cells. To test our hypothesis, we stimulated SVF cells isolated from WAT of wild-type mice with LPS. Indeed, LPS-treated SVF cells exhibited significantly higher *Osm* gene expression compared to vehicle-treated cells (Supplementary Fig. 3C). An even stronger LPS-

induced upregulation of *Osm* was found in RAW cells, a macrophage cell line (Supplementary Fig. 3D), rendering macrophages a plausible source of increased OSM production in WAT under obesity.

3.3 Inhibition of STAT3 activation prevents OSM-mediated reduction in Ucp1 expression

OSM was shown to inhibit pre-adipocyte differentiation (36) and to induce dedifferentiation of mature adipocytes (37), possibly providing an alternative explanation for the OSM-mediated reduction in UCP1 expression. Therefore, we assessed the amount of lipid droplets by means of Oil-Red-O staining and determined levels of the adipocyte differentiation marker PPARγ in cultured white adipocytes after OSM treatment. As depicted in Fig. 3A and Supplementary Fig. 4A-B, no changes in lipid droplet content and PPARγ protein and mRNA levels were observed in OSM-treated subcutaneous adipocytes compared to vehicle-treated control cells. Of note, in 3T3-L1 adipocytes PPARγ mRNA and protein levels were reduced after incubation with 100 ng/ml but not with lower doses of OSM (Supplementary Fig. 4C-F). Moreover, OSM treatment blunted insulin-induced phosphorylation of Akt in 3T3-L1 cells (Supplementary Fig. 4E and G), suggesting that OSM impacts on insulin signaling in white adipocytes. Collectively, these data indicate that a high OSM concentration may induce adipocyte dedifferentiation as previously described (37).

To elucidate whether ERK1/2 and/or STAT3 activation may contribute to OSM-mediated reduction in UCP1 expression, p-ERK1/2 and p-STAT3 abundance was assessed in mature subcutaneous adipocytes treated with OSM for different durations. As shown in Fig. 3A-C, OSM induced ERK1/2 phosphorylation after 15 minutes but continuously activated STAT3 at all time points over the course of 24 hours. To

independently study the contribution of the ERK and the STAT pathway to the action of OSM, we used the MEK inhibitor U0126 and the JAK inhibitor Tofacitinib, respectively. As expected, U0126 successfully prevented ERK1/2 phosphorylation while Tofacitinib completely blocked STAT3 activation (Fig. 3D). Pre-incubating subcutaneous adipocytes with U0126 had no effect on the OSM-mediated blunting of *Ucp1* mRNA levels (Fig. 3F). Conversely, Tofacitinib partially restored *Ucp1* mRNA expression (Fig. 3F). Thus, our data suggests that OSM suppresses browning in subcutaneous adipocytes JAK-STAT3 dependently.

3.4 OSM positively correlates with BMI but negatively with UCP1 in human subcutaneous WAT

To ascertain the clinical relevance of our mouse data, we measured *OSM* gene expression in subcutaneous fat biopsies from a cohort of lean and obese human subjects. A moderate but significant positive correlation between *OSM* mRNA levels and BMI was found (Fig. 4A). As expected, *IL-6* mRNA level also showed a similar positive correlation whereas *UCP1* gene expression negatively correlated with BMI (Fig. 4B, 4C). Furthermore, while no linear relationship between *UCP1* and *IL-6* gene expression levels was observed, *UCP1* negatively correlated with *OSM* expression (Fig. 4D, 4E). These results support our mouse data and indicate that elevated levels of OSM associated with obesity may suppress browning of WAT in human.

4. Discussion

Impaired WAT thermogenesis, i.e. the capacity of WAT to undergo browning, may contribute to the development and/or maintenance of obesity (38). Herein, we provide evidence that OSM may add to blunted WAT browning. In both mice and humans, increased adiposity is associated with a surge of proinflammatory cytokines including OSM in fat depots (20; 39). We simulated this *in vivo* observation by cultivating subcutaneous adipocytes in the presence of different OSM concentrations and observed a dose-dependent blunting of the main thermogenic marker UCP1. Conversely, expression of UCP1 and other browning markers was enhanced in ingWAT depots of HFD-fed gp130$^{\Delta adipo}$ mice with blocked OSM signaling. Clearly, we cannot exclude that other metabolic changes and/or other gp130 cytokines than OSM contributed directly or indirectly to the observed changes in ingWAT browning. Our human data confirms a previous small-scale study indicating that OSM is increased in subcutaneous WAT of people with obesity (20) and suggests that, in conjunction with our mouse data, obesity-induced OSM impairs WAT browning. In support of a physiological significance of such finding, *Ucp1* expression in WAT correlated negatively with body weight and BMI in mice and men, respectively, suggesting that elevated OSM levels contribute to increased body weight in both species. Of note, the gp130$^{\Delta adipo}$ mouse exhibits a complex phenotype that, besides direct effects on adipocytes, includes indirect metabolic changes in gut, pancreatic islets, liver and hypothalamus (12; 13; 28). In particular, knockout mice revealed reduced circulating levels of insulin and leptin, two factors that are critically involved in the regulation of energy expenditure and/or body weight (40; 41). Accordingly, such changes may have prevented a clearer effect on body weight and/or energy expenditure in these mice (12; 28). Moreover, when feeding mice a HFD, body weight

gain is heterogeneous, with some cohorts/litters being more resistant than others. In the current re-analysis, we included about twice as many litters compared to our initial analysis and found that the average body weight per litter was significantly reduced in HFD-fed knockout mice, supporting the notion that increased browning in knockout mice reduces body weight.

A more selective model to study OSM signaling *in vivo* is the adipocyte-specific OSM receptor knockout mouse ($OSMR^{\Delta adipo}$) (35). The OSM receptor dimerizes with a gp130 subunit to form the OSM receptor complex. Of note, the $OSMR^{\Delta adipo}$ mouse does not only block signaling of OSM but also of IL-31, as the latter also contains an OSMR subunit in its receptor complex (42; 43). Whether the $OSMR^{\Delta adipo}$ mouse also exhibits increased browning in ingWAT has not been reported. However, $OSMR^{\Delta adipo}$ mice are more sensitive to diet-induced obesity and display increased adipose tissue inflammation and insulin resistance (35). Hence, this phenotype clearly differs from the one of $gp130^{\Delta adipo}$ mice, which may be expected from the molecular difference between the two models. While OSM action is prevented in either model, it is accompanied by blocking of IL-31 signaling in the $OSMR^{\Delta adipo}$ mouse and via interfering with gp130 signaling in the $gp130^{\Delta adipo}$ mouse.

Using an *in vitro* model of WAT browning, we observed no direct effects of IL-6 and leptin on UCP1 expression. Considering that both IL-6 and leptin, like OSM, can signal via the JAK-STAT3 pathway, the question arises why OSM and not IL-6 or leptin mediates *Ucp1* transcription. One possibility is that their common negative regulator SOCS3 acts differently depending whether it is activated by OSM or by the other two factors. While SOCS3 was shown to prevent IL-6 mediated STAT3 phosphorylation after two hours (44), we observed herein that OSM promotes sustained activation of STAT3 up to 24 hours, suggesting a reduced inhibitory effect of SOCS3 at this OSM concentration. Other

explanations may include differential cross-talk with other cytokines (45) or differences in localization or composition of the receptors. It should be noted that, in contrast to OSM, IL-6 and leptin are generally considered positive regulators of WAT browning. For instance, more WAT browning was found after systemic IL-6 treatment (18) or in mouse models of cancer where IL-6 was upregulated under conditions of cachexia (19). Leptin likely promotes WAT browning in an indirect manner via signaling through the hypothalamus (30; 31). Since systemic leptin concentrations were lower in gp130$^{\Delta adipo}$ mice, we consider it improbable that leptin mediated the observed induction of ingWAT browning. Initially, we also hypothesized that the adipocyte-derived enzyme autotaxin may have contributed to elevated browning in gp130$^{\Delta adipo}$ mice, especially since an obesity-associated increase in gp130 signaling upregulated autotaxin in adipose tissue (34) and mice that transgenically overexpressed autotaxin displayed reduced browning in WAT (33). Yet, the observation that autotaxin levels in ingWAT were not different between HFD-fed gp130$^{\Delta adipo}$ and gp130$^{F/F}$ mice infers an insignificant role of autotaxin on the browning phenotype in our mouse model.

The present study confirms previous work (20; 29) showing that *Osm* levels are elevated in adipose tissue of obese mice, likely due to an altered gene expression in the non-adipocyte fraction. We further find that an LPS-mediated inflammatory response induced *Osm* gene expression in SVF cells as well as cultured macrophages, providing one potential mechanism driving such increase. Despite the putatively higher *Osm* level in all fat depots, the unchanged UCP1 expression in BAT of HFD-fed gp130$^{\Delta adipo}$ mice would infer that the inhibitory role of OSM is limited to ingWAT. This notion is underpinned by a recent study by Sanchez-Infantes *et al.*, who found that cold-induced upregulation of *Ucp1* was blunted in ingWAT of mice after local OSM infusion while no such effect was

observed when OSM was infused in supraclavicular BAT (29). Besides higher OSM levels, increased OSM signaling may be effectuated by elevated OSM sensitivity or augmented expression of gp130 and/or OSM receptor (OSMR). However, while OSM was shown to induce *gp130* and *Osmr* levels in 3T3-L1 cells (35), we did not find increased *gp130* mRNA levels in adipocytes of HFD compared to chow-fed mice. Collectively, these data suggest that lowering of local OSM secretion and/or signaling in WAT could be a potential therapeutic strategy to promote browning and, thus, to combat obesity. In contrast, a recent study reported that systemic treatment with OSM improved the metabolic phenotype of obese mice (46). This underscores the importance to understand the differential role of OSM in various tissues.

In the current study, we employed Tofacinib to block OSM-mediated STAT3 signaling in cultured adipocytes. This novel JAK2 inhibitor successfully induced *UCP1* mRNA in human adipocytes (16) and promoted WAT browning in HFD-fed mice (47). However, herein we did not observe induction of *Ucp1* by Tofacitinib in cultured adipocytes. Potentially, isoprotenerol may have masked such effect of Tofacitinib. Nonetheless, our findings are in agreement with the conception that JAK2 inhibition in white adipocytes may be beneficial in obese conditions.

In conclusion, our data support the notion that OSM negatively regulates thermogenesis in subcutaneous WAT via gp130-STAT3 signaling and, thus, may be an attractive target to treat obesity.

Acknowledgments

This work was supported by a grant from the Swiss National Science Foundation (#310030-179344 to DK), a grant from the Wolfermann-Nägeli Stiftung and a grant from the Hartmann-Müller Stiftung, University of Zurich (#2292) (both to SW). Human studies were supported by the Deutsche Forschungsgemeinschaft (DFG, German Research Foundation) through CRC 1052, project number 209933838, subproject B1 to MBl. We are grateful to Werner Muller, Faculty of Biology, Medicine and Health, University of Manchester, U.K., for providing gp130$^{\Delta adipo}$ mice.

Author's contribution

P.P.v.K and S.W. designed and performed experiments, analyzed data and wrote the manuscript. T.S.O., M.Bo and M.Bl. performed experiments. D.K. designed experiments, analyzed data and wrote the manuscript. All authors reviewed and commented on the manuscript.

Declaration of Interests

The authors declare no competing interests.

Appendix A: Supplementary Data

References

1. Cannon B, Nedergaard J: Brown adipose tissue: function and physiological significance. Physiol Rev 2004;84:277-359
2. Kajimura S, Spiegelman BM, Seale P: Brown and Beige Fat: Physiological Roles beyond Heat Generation. Cell Metab 2015;22:546-559
3. Cypess AM, Lehman S, Williams G, Tal I, Rodman D, Goldfine AB, Kuo FC, Palmer EL, Tseng YH, Doria A, Kolodny GM, Kahn CR: Identification and importance of brown adipose tissue in adult humans. N Engl J Med 2009;360:1509-1517
4. van Marken Lichtenbelt WD, Vanhommerig JW, Smulders NM, Drossaerts JM, Kemerink GJ, Bouvy ND, Schrauwen P, Teule GJ: Cold-activated brown adipose tissue in healthy men. N Engl J Med 2009;360:1500-1508
5. Virtanen KA, Lidell ME, Orava J, Heglind M, Westergren R, Niemi T, Taittonen M, Laine J, Savisto NJ, Enerback S, Nuutila P: Functional brown adipose tissue in healthy adults. N Engl J Med 2009;360:1518-1525
6. Bartelt A, Heeren J: Adipose tissue browning and metabolic health. Nat Rev Endocrinol 2014;10:24-36
7. Giordano A, Frontini A, Cinti S: Convertible visceral fat as a therapeutic target to curb obesity. Nat Rev Drug Discov 2016;15:405-424
8. Donath MY, Shoelson SE: Type 2 diabetes as an inflammatory disease. Nat Rev Immunol 2011;11:98-107
9. Ma D, Wang Y, Zhou G, Wang Y, Li X: Review: the Roles and Mechanisms of Glycoprotein 130 Cytokines in the Regulation of Adipocyte Biological Function. Inflammation 2019;42:790-798
10. Taga T, Kishimoto T: Gp130 and the interleukin-6 family of cytokines. Annu Rev Immunol 1997;15:797-819
11. White UA, Stewart WC, Stephens JM: Gp130 cytokines exert differential patterns of crosstalk in adipocytes both in vitro and in vivo. Obesity (Silver Spring) 2011;19:903-910
12. Wueest S, Item F, Lucchini FC, Challa TD, Muller W, Bluher M, Konrad D: Mesenteric Fat Lipolysis Mediates Obesity-Associated Hepatic Steatosis and Insulin Resistance. Diabetes 2016;65:140-148
13. Wueest S, Laesser CI, Boni-Schnetzler M, Item F, Lucchini FC, Borsigova M, Muller W, Donath MY, Konrad D: IL-6-Type Cytokine Signaling in Adipocytes Induces Intestinal GLP-1 Secretion. Diabetes 2018;67:36-45
14. Babaei R, Schuster M, Meln I, Lerch S, Ghandour RA, Pisani DF, Bayindir-Buchhalter I, Marx J, Wu S, Schoiswohl G, Billeter AT, Krunic D, Mauer J, Lee YH, Granneman JG, Fischer L, Muller-Stich BP, Amri EZ, Kershaw EE, Heikenwalder M, Herzig S, Vegiopoulos A: Jak-TGFbeta cross-talk links transient adipose tissue inflammation to beige adipogenesis. Sci Signal 2018;11
15. Banks AS, McAllister FE, Camporez JP, Zushin PJ, Jurczak MJ, Laznik-Bogoslavski D, Shulman GI, Gygi SP, Spiegelman BM: An ERK/Cdk5 axis controls the diabetogenic actions of PPARgamma. Nature 2015;517:391-395
16. Moisan A, Lee YK, Zhang JD, Hudak CS, Meyer CA, Prummer M, Zoffmann S, Truong HH, Ebeling M, Kiialainen A, Gerard R, Xia F, Schinzel RT, Amrein KE, Cowan CA: White-to-brown metabolic conversion of human adipocytes by JAK inhibition. Nat Cell Biol 2015;17:57-67

17. Zhang Y, Li R, Meng Y, Li S, Donelan W, Zhao Y, Qi L, Zhang M, Wang X, Cui T, Yang LJ, Tang D: Irisin stimulates browning of white adipocytes through mitogen-activated protein kinase p38 MAP kinase and ERK MAP kinase signaling. Diabetes 2014;63:514-525
18. Knudsen JG, Murholm M, Carey AL, Bienso RS, Basse AL, Allen TL, Hidalgo J, Kingwell BA, Febbraio MA, Hansen JB, Pilegaard H: Role of IL-6 in exercise training- and cold-induced UCP1 expression in subcutaneous white adipose tissue. PLoS One 2014;9:e84910
19. Petruzzelli M, Schweiger M, Schreiber R, Campos-Olivas R, Tsoli M, Allen J, Swarbrick M, Rose-John S, Rincon M, Robertson G, Zechner R, Wagner EF: A switch from white to brown fat increases energy expenditure in cancer-associated cachexia. Cell Metab 2014;20:433-447
20. Sanchez-Infantes D, White UA, Elks CM, Morrison RF, Gimble JM, Considine RV, Ferrante AW, Ravussin E, Stephens JM: Oncostatin m is produced in adipose tissue and is regulated in conditions of obesity and type 2 diabetes. J Clin Endocrinol Metab 2014;99:E217-225
21. Lam YY, Ha CW, Campbell CR, Mitchell AJ, Dinudom A, Oscarsson J, Cook DI, Hunt NH, Caterson ID, Holmes AJ, Storlien LH: Increased gut permeability and microbiota change associate with mesenteric fat inflammation and metabolic dysfunction in diet-induced obese mice. PLoS One 2012;7:e34233
22. Rolle-Kampczyk U, Gebauer S, Haange SB, Schubert K, Kern M, Moulla Y, Dietrich A, Schon MR, Kloting N, von Bergen M, Bluher M: Accumulation of distinct persistent organic pollutants is associated with adipose tissue inflammation. Sci Total Environ 2020;748:142458
23. Betz UA, Bloch W, van den Broek M, Yoshida K, Taga T, Kishimoto T, Addicks K, Rajewsky K, Muller W: Postnatally induced inactivation of gp130 in mice results in neurological, cardiac, hematopoietic, immunological, hepatic, and pulmonary defects. J Exp Med 1998;188:1955-1965
24. Kovsan J, Osnis A, Maissel A, Mazor L, Tarnovscki T, Hollander L, Ovadia S, Meier B, Klein J, Bashan N, Rudich A: Depot-specific adipocyte cell lines reveal differential drug-induced responses of white adipocytes--relevance for partial lipodystrophy. Am J Physiol Endocrinol Metab 2009;296:E315-322
25. Rapold RA, Wueest S, Knoepfel A, Schoenle EJ, Konrad D: Fas activates lipolysis in a Ca2+-CaMKII-dependent manner in 3T3-L1 adipocytes. J Lipid Res 2013;54:63-70
26. Wueest S, Mueller R, Bluher M, Item F, Chin AS, Wiedemann MS, Takizawa H, Kovtonyuk L, Chervonsky AV, Schoenle EJ, Manz MG, Konrad D: Fas (CD95) expression in myeloid cells promotes obesity-induced muscle insulin resistance. EMBO Mol Med 2014;6:43-56
27. Xu Y, Wang Y, Liu J, Cao W, Li L, Du H, Zhan E, Zhang R, Liu H, Xu M, Chen T, Qu Y, Yu B: Adipose tissue-derived autotaxin causes cardiomyopathy in obese mice. J Mol Endocrinol 2019;63:113-121
28. Odermatt TS, Dedual MA, Borsigova M, Wueest S, Konrad D: Adipocyte-specific gp130 signalling mediates exercise-induced weight reduction. Int J Obes (Lond) 2020;44:707-714
29. Sanchez-Infantes D, Cereijo R, Peyrou M, Piquer-Garcia I, Stephens JM, Villarroya F: Oncostatin m impairs brown adipose tissue thermogenic function and the browning of subcutaneous white adipose tissue. Obesity (Silver Spring) 2017;25:85-93

30. Dodd GT, Decherf S, Loh K, Simonds SE, Wiede F, Balland E, Merry TL, Munzberg H, Zhang ZY, Kahn BB, Neel BG, Bence KK, Andrews ZB, Cowley MA, Tiganis T: Leptin and insulin act on POMC neurons to promote the browning of white fat. Cell 2015;160:88-104
31. Plum L, Rother E, Munzberg H, Wunderlich FT, Morgan DA, Hampel B, Shanabrough M, Janoschek R, Konner AC, Alber J, Suzuki A, Krone W, Horvath TL, Rahmouni K, Bruning JC: Enhanced leptin-stimulated Pi3k activation in the CNS promotes white adipose tissue transdifferentiation. Cell Metab 2007;6:431-445
32. Rodriguez A, Becerril S, Mendez-Gimenez L, Ramirez B, Sainz N, Catalan V, Gomez-Ambrosi J, Fruhbeck G: Leptin administration activates irisin-induced myogenesis via nitric oxide-dependent mechanisms, but reduces its effect on subcutaneous fat browning in mice. Int J Obes (Lond) 2015;39:397-407
33. Federico L, Ren H, Mueller PA, Wu T, Liu S, Popovic J, Blalock EM, Sunkara M, Ovaa H, Albers HM, Mills GB, Morris AJ, Smyth SS: Autotaxin and its product lysophosphatidic acid suppress brown adipose differentiation and promote diet-induced obesity in mice. Mol Endocrinol 2012;26:786-797
34. Sun S, Wang R, Song J, Guan M, Li N, Zhang X, Zhao Z, Zhang J: Blocking gp130 signaling suppresses autotaxin expression in adipocytes and improves insulin sensitivity in diet-induced obesity. J Lipid Res 2017;58:2102-2113
35. Elks CM, Zhao P, Grant RW, Hang H, Bailey JL, Burk DH, McNulty MA, Mynatt RL, Stephens JM: Loss of Oncostatin M Signaling in Adipocytes Induces Insulin Resistance and Adipose Tissue Inflammation in Vivo. J Biol Chem 2016;291:17066-17076
36. Miyaoka Y, Tanaka M, Naiki T, Miyajima A: Oncostatin M inhibits adipogenesis through the RAS/ERK and STAT5 signaling pathways. J Biol Chem 2006;281:37913-37920
37. Song HY, Kim MR, Lee MJ, Jeon ES, Bae YC, Jung JS, Kim JH: Oncostatin M decreases adiponectin expression and induces dedifferentiation of adipocytes by JAK3- and MEK-dependent pathways. Int J Biochem Cell Biol 2007;39:439-449
38. Rui L: Brown and Beige Adipose Tissues in Health and Disease. Compr Physiol 2017;7:1281-1306
39. Komori T, Tanaka M, Senba E, Miyajima A, Morikawa Y: Deficiency of oncostatin M receptor beta (OSMRbeta) exacerbates high-fat diet-induced obesity and related metabolic disorders in mice. J Biol Chem 2014;289:13821-13837
40. Pandit R, Beerens S, Adan RAH: Role of leptin in energy expenditure: the hypothalamic perspective. Am J Physiol Regul Integr Comp Physiol 2017;312:R938-R947
41. Porte D, Jr., Baskin DG, Schwartz MW: Insulin signaling in the central nervous system: a critical role in metabolic homeostasis and disease from C. elegans to humans. Diabetes 2005;54:1264-1276
42. Cornelissen C, Luscher-Firzlaff J, Baron JM, Luscher B: Signaling by IL-31 and functional consequences. Eur J Cell Biol 2012;91:552-566
43. Dillon SR, Sprecher C, Hammond A, Bilsborough J, Rosenfeld-Franklin M, Presnell SR, Haugen HS, Maurer M, Harder B, Johnston J, Bort S, Mudri S, Kuijper JL, Bukowski T, Shea P, Dong DL, Dasovich M, Grant FJ, Lockwood L, Levin SD, LeCiel C, Waggie K, Day H, Topouzis S, Kramer J, Kuestner R, Chen Z, Foster D, Parrish-Novak J, Gross JA: Interleukin 31, a cytokine produced by activated T cells, induces dermatitis in mice. Nat Immunol 2004;5:752-760

44. Wunderlich CM, Hovelmeyer N, Wunderlich FT: Mechanisms of chronic JAK-STAT3-SOCS3 signaling in obesity. JAKSTAT 2013;2:e23878
45. Zvonic S, Baugh JE, Jr., Arbour-Reily P, Mynatt RL, Stephens JM: Cross-talk among gp130 cytokines in adipocytes. J Biol Chem 2005;280:33856-33863
46. Komori T, Tanaka M, Furuta H, Akamizu T, Miyajima A, Morikawa Y: Oncostatin M is a potential agent for the treatment of obesity and related metabolic disorders: a study in mice. Diabetologia 2015;58:1868-1876
47. Qurania KR, Ikeda K, Wardhana DA, Barinda AJ, Nugroho DB, Kuribayashi Y, Rahardini EP, Rinastiti P, Ryanto GRT, Yagi K, Hirata KI, Emoto N: Systemic inhibition of Janus kinase induces browning of white adipose tissue and ameliorates obesity-related metabolic disorders. Biochem Biophys Res Commun 2018;502:123-128

Figure Legends

Figure 1 Increased browning in ingWAT of HFD-fed gp130$^{\Delta adipo}$ mice

(A) Representative Western blot of respective targets in ingWAT from HFD-fed gp130$^{F/F}$ and gp130$^{\Delta adipo}$ mice. (B) Quantification of UCP1 protein levels in ingWAT of HFD-fed gp130$^{F/F}$ (n=11) and gp130$^{\Delta adipo}$ (n=13) mice. (C) Gene expression levels of respective targets in ingWAT from HFD-fed gp130$^{F/F}$ (n=8-11) and gp130$^{\Delta adipo}$ mice (n=9-14). (D) Body weight of HFD-fed gp130$^{F/F}$ and gp130$^{\Delta adipo}$ mice at time of sacrifice and corresponding *Ucp1* gene expression level in ingWAT (n=22). (E-F) UCP1 and corresponding p-ERK1/2 or p-STAT3 protein levels in ingWAT from HFD-fed gp130$^{F/F}$ and gp130$^{\Delta adipo}$ mice (n=21). Open circles/squares represent individual gp130$^{F/F}$ and gp130$^{\Delta adipo}$ ingWAT samples, respectively. *, $p<0.05$; **, $p<0.01$. Panel B, Mann-Whitney test; panel C, Student's t-test; panel D, E and F, Linear Regression analysis. Error bars represent S.E.

Figure 2 OSM dose-dependently lowers isoproterenol-induced UCP1 levels in cultured adipocytes

Normalized *Ucp1* gene expression and/or UCP1 protein levels of subcutaneous adipocytes (A-B) or 3T3-L1 adipocytes (C) cultured in the presence of a vehicle or different concentrations of OSM or IL-6 for 24 hours followed by 6 hours of isoproterenol stimulation (n=4-8). **, $p<0.01$; ***, $p<0.001$ compared to vehicle-treated control. Panel A and B, One-way ANOVA. Error bars represent S.E.

Figure 3 Inhibition of STAT3 activation prevents OSM-mediated reduction in Ucp1 expression

(A-C) Western blot of subcutaneous white adipocytes stimulated with OSM or vehicle for indicated time periods and quantifications of phosphorylated ERK1/2 and STAT3 protein levels (n=4). (D) Western blot of subcutaneous white adipocytes stimulated with vehicle (control), U0126 (50 μM) or Tofacitinib (2 μM) for 1 hour. (E-F) *Ucp1* gene expression levels of subcutaneous white adipocytes that were subsequently stimulated with either U0126 or Tofacitinib (1 hour), OSM (3 hours) and isoproterenol (3 hours), displayed relative to a negative control (left panel) or as a percentage of the internal control with and without inhibitor (right panel; n=6-8). *, $p<0.05$; **, $p<0.01$; ***, $p<0.001$. One-way ANOVA (E-F, left panel) or Student's t-test (E-F, right panel). Error bars represent S.E.

Figure 4 OSM positively correlates with BMI but negatively with UCP1 in human subcutaneous WAT

(A-E) Correlations between BMI and *OSM*, *IL-6* or *UCP1* mRNA level, and between *UCP1* and *OSM* or *IL-6* mRNA levels (n=66). Gene expression data was obtained from subcutaneous human WAT biopsies. Samples with undetectable gene expression levels were excluded from analysis.

Figure 1

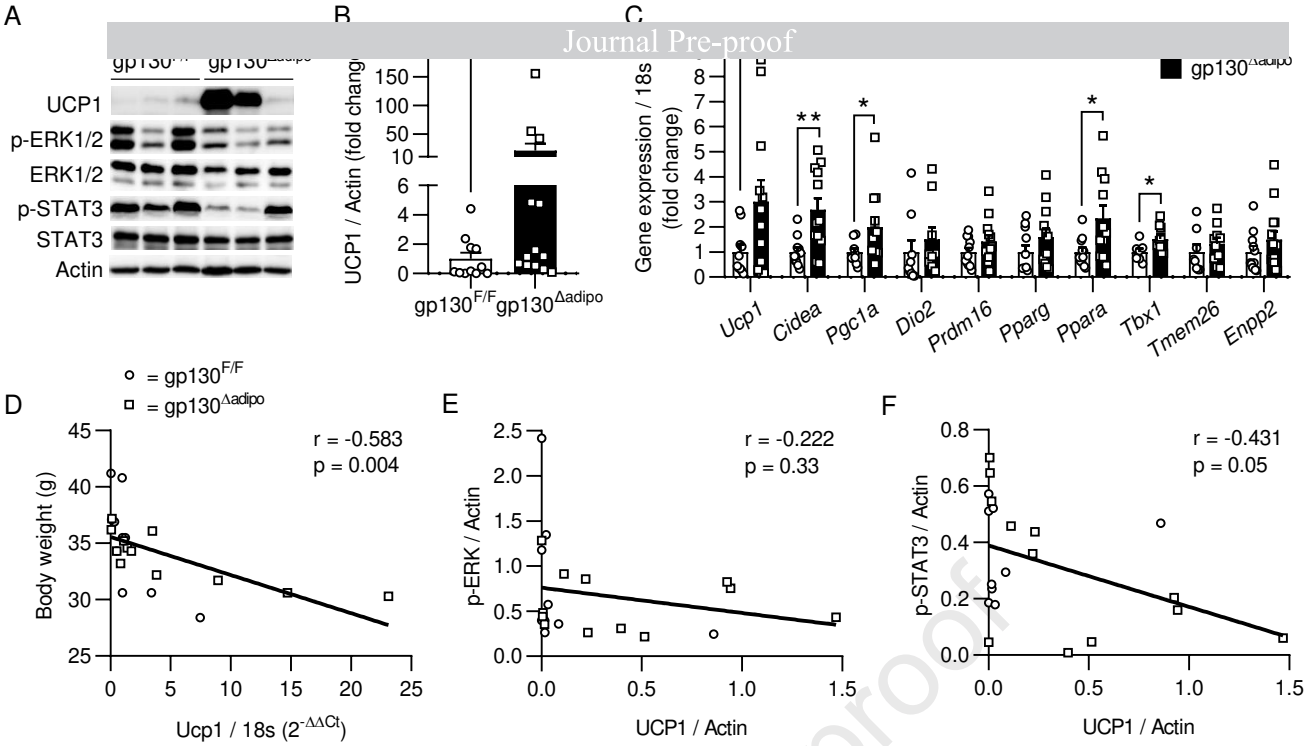


Figure 2

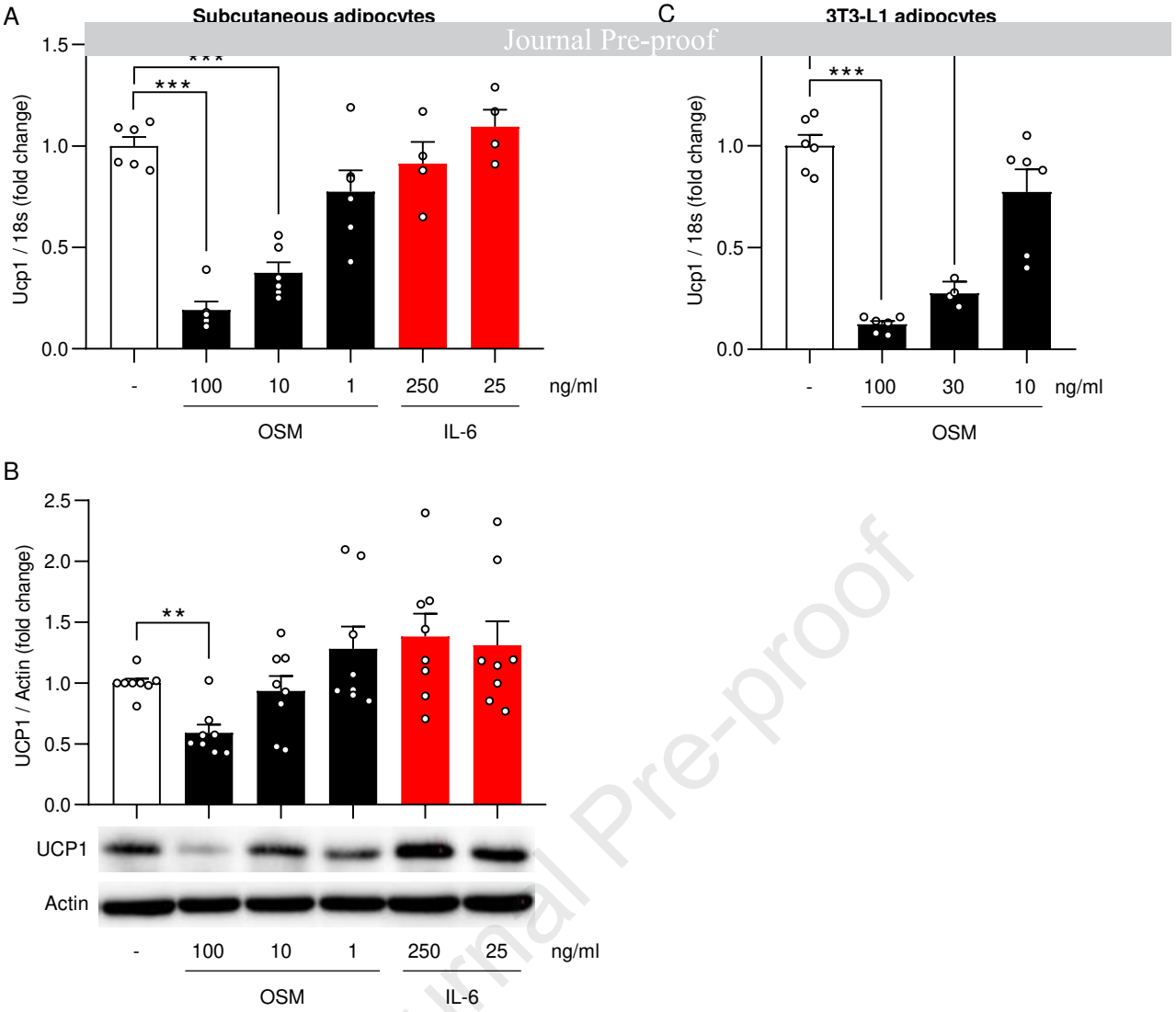


Figure 3

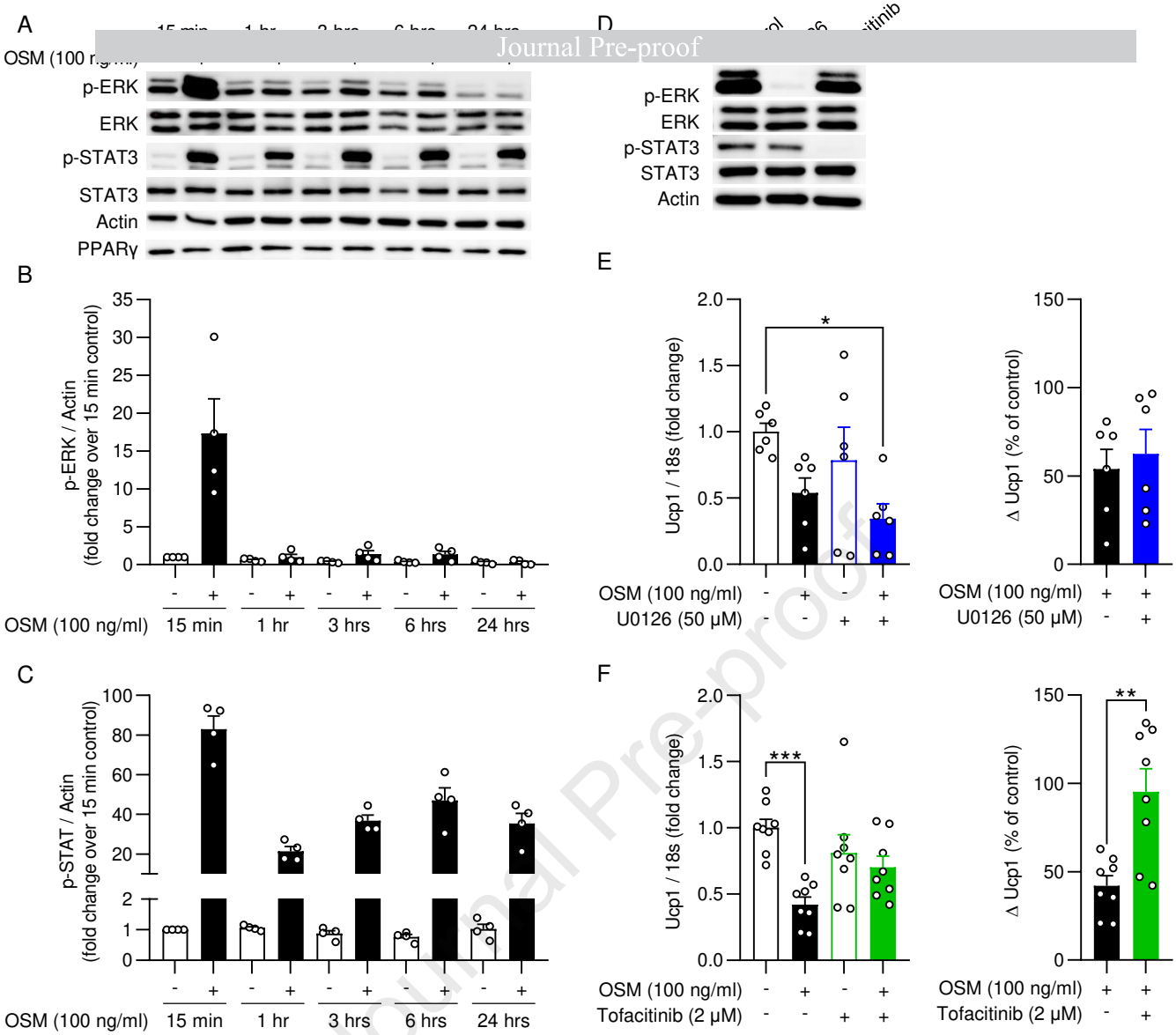
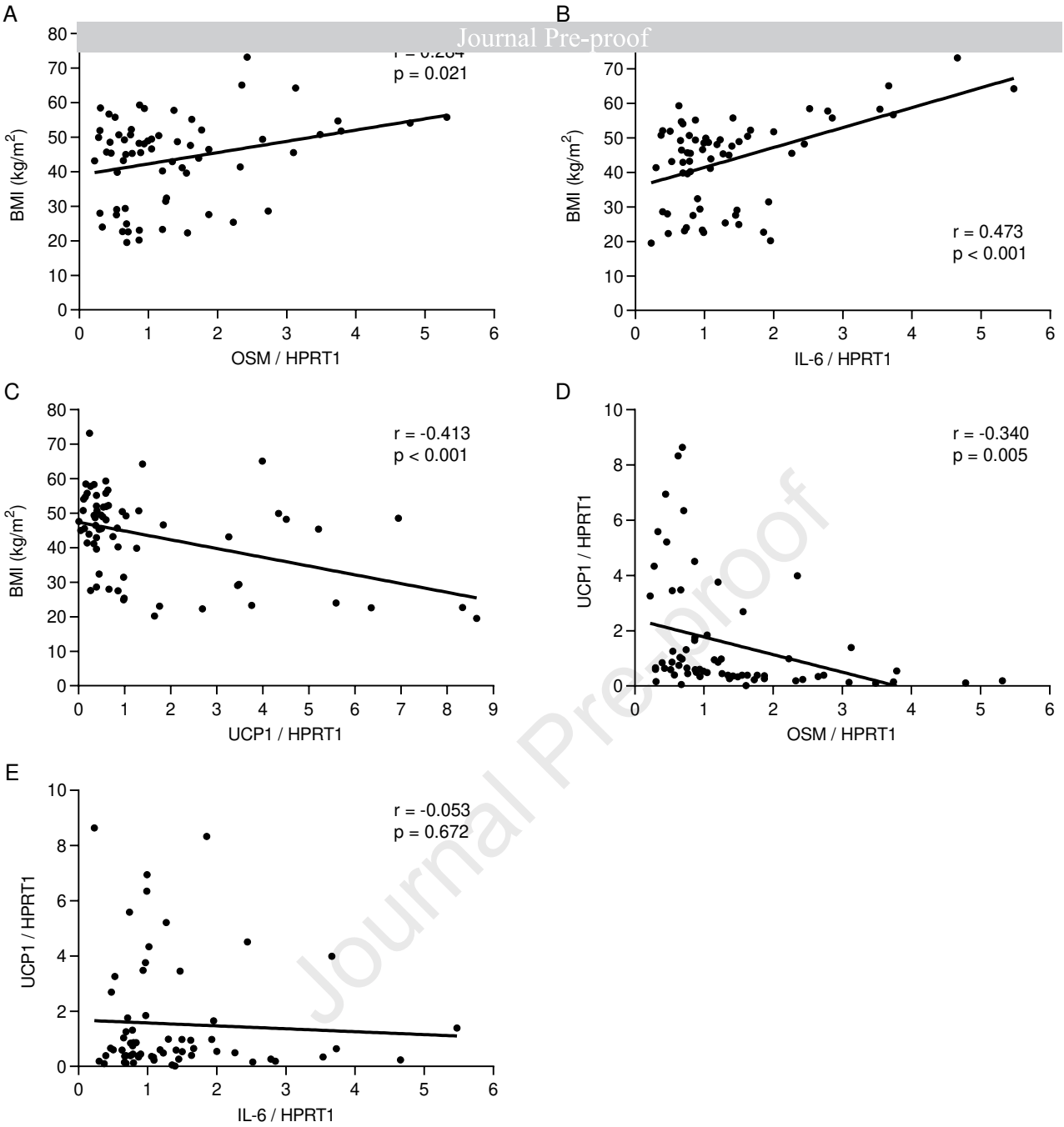


Figure 4



Highlights

- OSM is regulated under obesity and negatively correlates with UCP1 in WAT
- OSM suppresses isoproterenol-induced UCP1 in subcutaneous adipocytes
- OSM signals through the gp130-STAT3 pathway to lower UCP1 expression
- Obese mice lacking gp130 in adipocytes exhibit increased WAT browning

Declaration of Interests

The authors declare no competing interests.